## Tissue DNA Extraction Kit Cat. No. A101211

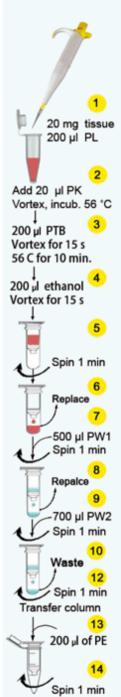
## **Before starting**

- 1. Add 10 ml of absolute ethanol to the PW1 (only at the first use).
- 2. Add 48 ml of absolute ethanol to the PW2 (only at the first use).
- 3. Add Proteinase K (PK) solution to the lyophilized powder of proteinase K and store at -20 °C until usage (only at the first use).
- Check PW1, PL and PTB for salt precipitation. Redissolve any precipitation at 50 ℃.
- 5. Preheat the solution of PE to 56 °C before starting the extraction process to enhance DNA extraction yield.

## **Tissue DNA Extraction Protocol**

- 1. Transfer 20 mg of tissue (10 mg for liver or spleen) to a 1.5 ml tube and add 200 µl of PL solution. Cutting the tissue into the small pieces increases the yield of genomic DNA and reduce lysis incubation time.
- 2. Add 20  $\mu$ l of Proteinase K and mix them well by vortexing and incubate at 56 °C until complete lysis (vortex occasionally). Lysis time varies depending on the tissue type.
- 3. After lysis of tissue, add 200 µl of PTB solution and vortex for 15 seconds and incubate at 56 °C for 10 minutes.
- 4. Add 200 µl of absolute ethanol and mix by puls-vortxing (15 s).
- Carefully trasfer lysate to the spin column. A quick spin before lysate transfer would be prefered if there was any debries in the mixture. Spin column for 1 min at 13,000 rpm.
- 6. Replace the collection tube with a new one.
- 7. Add 500 µl of PW1 into the column and spin for 1 min at 13,000 rpm.
- 8. Replace the collection tube with a new one.
- 9. Add 700 µl of PW2 into the column and spin for 1 min at 13,000 rpm.
- Pour off the flow-through of collection tube.
- 11. Repeat step 8 and 9 with 500 µl of PW2 (optional)
- 12. Spin for 1 min at 13,000 rpm to remove the remaining of the wash buffer. Transfer the spin column to a new 1.5 ml microtube.
- 13. Add 200 µl of preheated PE, wait 3 min at room temprature. If you want more concentration add less PE (100 µl).
- 14. Spin for 1 min at 13,000 rpm to elute DNA from the column. Store DNA solution at -20 °C.

Tel: +98 (511) 8551360-2



Fax: +98 (511) 8551363