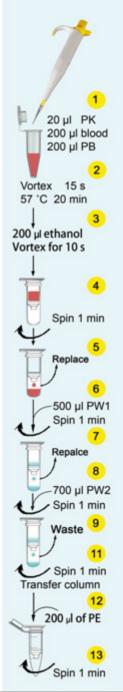


Before starting

- 1. Add 10 ml of absolute ethanol to the PW1 (only at the first use).
- 2. Add 48 ml of absolute ethanol to the PW2 (only at the first use).
- 3. Add Proteinase K (PK) solution to the lyophilized powder of proteinase K and store at -20 °C until usage (only at the first use).
- Check PW1 and PB for salt precipitation. Redissolve any preciptation at 50 ℃.
- 5. Preheat the solution of PE to 56 °C before starting the extraction process to enhance DNA extraction yield.

Blood DNA Extraction Protocol

- 1. Add 20 μ l of proteinase K, 200 μ l of blood and finally 200 μ l of PB into a 1.5 ml microtube.
- 2. Mix them well by vortexing (15 s) and incubate at 57 °C for 20 min.
- 3. Add 200 µl of absolute ethanol and mix it by vortexing (10 s).
- 4. After a quick spin, carefully trasfer lysate to the spin column. Do not touch upper rim of column. Spin for 1 min at 13,000 rpm. If you see blood on the column, repeat the spin for 1 min.
- 5. Replace the collection tube with a new one.
- 6. Add 500 µl of PW1 and spin for 1 min at 13,000 rpm.
- 7. Replace the collection tube with a new one.
- 8. Add 700 µl of PW2 and spin for 1 min at 13,000 rpm.
- Pour off the flow-through of collection tube.
- 10. Repeat step 8 and 9 with 500 µl of PW2 (optional)
- 11. Spin for 1 min at 13,000 rpm to remove the remaining of the wash buffer. Transfer the spin column to a new 1.5 ml microtube.
- 12. Add 200 μ l of preheated PE, wait 3 min at room temprature. If you want more concentration add less PE (100 μ l).
- 13. Spin for 1 min at 13,000 rpm to elute DNA from the column. Store DNA solution at -20 °C.



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