



Before Starting

1. Add 48 ml of absolute ethanol to the PW (only at the first use).
2. Check PW, GB for salt precipitation. Redissolve any precipitation at 50 °C.
3. Preheat the solution of PE to 56 °C before starting the extraction process to enhance DNA extraction yield.

Reagents NOT provided

1. Isopropanol
2. Absolute ethanol

Protocol

1. Cutting the agarose gel, as small as possible and weight the gel slice in a clean microtube. Add three volumes of gel binding buffer (GB). If you want to purify a PCR product or restriction enzyme reaction, add 5 volumes of GB to the solution. if your solution volume is less than 50 μ l, consider it 50 μ l and add 250 μ l GB to the mixture.
2. Incubate at 60 °C for 15 min and vortex the tube every 2-3 min, until complete resolving of gel (DNA extraction from solutions do not need this step).
3. Add one volume of isopropanol to the mixture.
4. Transfer mixture to the column and spin for 2 min at 8,000 rpm.
5. Pour off the flow-through of collection tube.
6. Add 700 μ l of PW and spin for 1 min at 13,000 rpm.
7. Pour off the flow-through of collection tube.
8. Repeat step 6 and 7 with 500 μ l of PW (optional)
9. Spin the column for 2 min at 13,000 rpm to remove the remaining of the wash buffer. Transfer the spin column to a new 1.5 ml microtube.
10. Add 50 μ l of pre-warmed PE, wait 3 min at room temperature. If you want more concentration add less pre-warmed PE (30 μ l).
11. Spin for 1 min at 13,000 rpm to elute DNA from the column. Store DNA solution at -20 °C.

